

Nucleophilic Addition of Potassium *O*-Ethyl Dithiocarbonate to Baylis-Hillman Adducts Using 9-BBN as Catalyst

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Received May 11, 2009; accepted June 26, 2009

Abstract. An efficient diastereoselective Michael addition of the commercial potassium *O*-ethyl dithiocarbonate to Baylis-Hillman adducts in the presence of 9-BBN (9-borabicyclo[3.3.1]nonane) as a Lewis acid is reported. 9-BBN both protected the hydroxyl group and activated the carbonyl of the Michael acceptor.

Keywords: Xanthates; Baylis-Hillman adducts; 9-BBN as a Lewis acid.

Resumen. En este artículo se describe una adición de Michael diastereoselectiva del *O*-etil ditiocarbonato de potasio comercial sobre aductos de Baylis-Hillman, utilizando 9-BBN (9-borabicyclo[3.3.1]nonano) como ácido de Lewis. El 9-BBN protegió el grupo hidroxilo y activó al mismo tiempo el grupo carbonilo del aceptor de Michael.

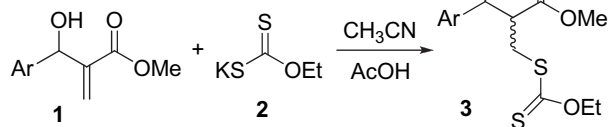
Keywords: Xantatos; aductos de Baylis-Hillman; 9-BBN, ácido de Lewis.

Introduction

The densely functionalized Baylis-Hillman adducts have considerable preparative value in the synthesis of complex molecules [1]. Most Michael addition processes on Baylis-Hillman adducts involve the loss of the hydroxyl group, an important functional group in the adduct structure. Except for a few documented cases [2], the loss of this group is typically prevented by incorporating an undesirable extra alcohol-protection/deprotection sequence into the synthetic plan [3]. In recent years xanthates (dithiocarbonates)[4] have found widespread applications in organic syntheses of complex structures and in living radical polymerization chemistry, mainly exploiting the efficiency of these compounds in the production of alkyl free radicals [5].

Results and discussion

In recent efforts to synthesize the xanthate **3**, we examined the nucleophilic addition of the commercial potassium *O*-ethyl dithiocarbonate to a Baylis-Hillman adduct without the loss of the hydroxyl group (Scheme 1). Specifically, we examined the reaction of the xanthate salt with the Baylis-Hillman adduct **1** in acetonitrile and acetic acid (as a proton source), stirring the reaction mixture at different temperatures. However, after 24 h of reaction, most of the starting hydroxyester **1** was recovered unchanged and a 1:1 diastereoisomeric mixture of the desired xanthate **3** was isolated in rather low yields (~10%).



Scheme 1.

Looking to improve the process, we realized that the hydroxyl function in the structure of the Baylis-Hillman adduct is in a suitable position to hold a Lewis acid, which would automatically activate the carbonyl function in a six-member structure. Thus, it was hypothesized that a borane, specifically a one-hydride transfer borane such as 9-BBN [6], could be useful not only to activate the Michael acceptor but also to protect the hydroxyl group. Thus the first step in the process might be an acid-base reaction between the hydride and the proton of the hydroxyl group, leading to the formation of the complex **4** (Figure 1).

To confirm this hypothesis, several Baylis-Hillman adducts were prepared and subjected to reaction with potassium *O*-ethyl dithiocarbonate (2 equiv), 9-BBN (1.5 equiv), and acetic acid (1 equiv), in acetonitrile at room temperature. Under these conditions, the desired xanthate was obtained generally in good yields (Table 1). Interestingly, we found that the reaction proceeded with some degree of diastereoselectivity, with the *anti* isomer being the major product at least in the case of **4b**. The relative configuration of this latter product was confirmed by X-ray crystallography of the *t*-butyldimethylsilyl derivative of the major product of **4b** (Figure 2) [7].

This stereochemical outcome might be rationalized in terms of the two possible intermediates depicted in scheme 2. If we

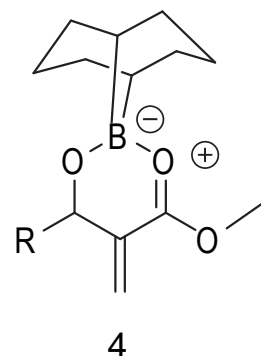
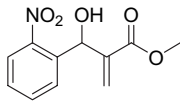
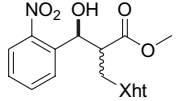
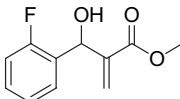
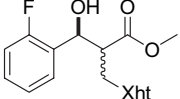
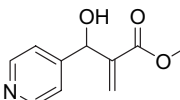
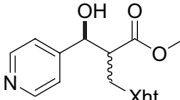
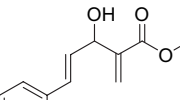
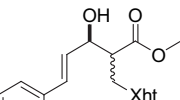
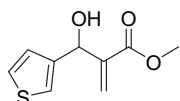
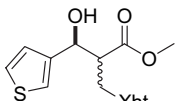
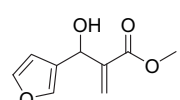
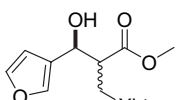
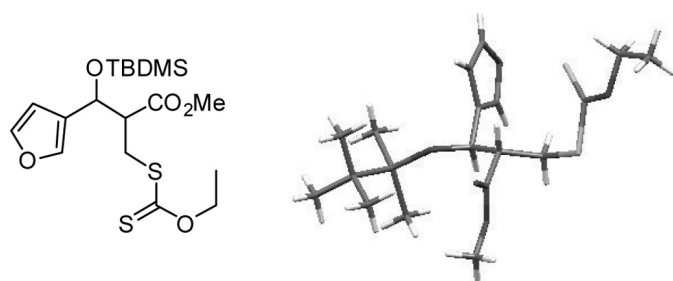


Fig. 1.

Table 1. Addition of potassium *O*-ethyl dithiocarbonate *O*-ethylxanthate to Baylis-Hillman adducts.

Baylis-Hillman	Compound	Product	Yield %	dr anti-syn
	4a		82	78:22
	4b		87	85:15
	4c		71	71:29
	4d		74	N.D.
	4e		95	81:19
	4f		57	66:34

Xht = SCSOC₂H₅. N.D. is not determined.

**Fig. 2.** X-Ray crystallography of the TBDMS-protected major isomer of 4b.

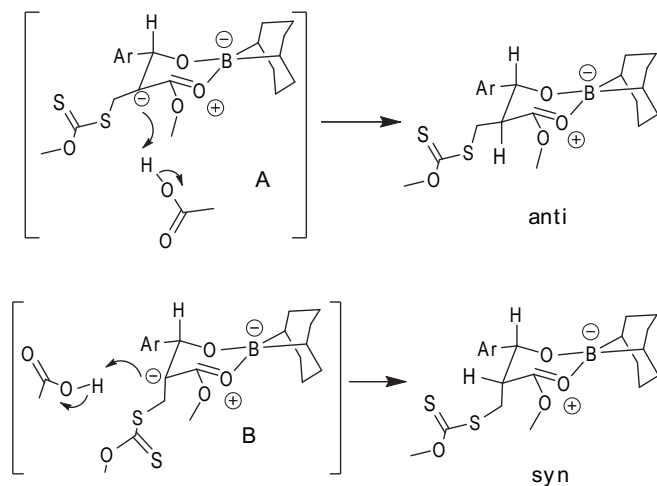
supposed that the key intermediate generates a six-membered ring, the preferred chair configuration provides the larger functional groups in equatorial position **A**, avoiding the axial interactions present in the **B** intermediate in which the bulky methylenxanthate group is in an axial position. The former enolate **A** leads to the incoming protonation in the axial position anti with respect to the adjacent hydrogen in the ring. Thus, 9-BBN can play an indispensable role in the stereocontrol of the addition reaction.

Conclusion

In closing, an efficient new methodology for synthesizing xanthates via 1,4-addition of xanthates to Baylis-Hillman adducts, promoted by 9-BBN as a Lewis acid with control of the diastereoselectivity, is reported. 9-BBN both protects the hydroxyl group of the Baylis-Hillman adduct and activates the carbonyl group of the Michael acceptor.

Experimental Section

For all reactions, a ~0.5 M 9-BBN (Aldrich) solution in tetrahydrofuran was used. All column chromatography purifications were carried out using silica gel (230-400 mesh ASTM; Merck). TLC was performed on precoated aluminum-backed plates with silica gel 60 F₂₅₄ (2 mm; Merck), which were developed using standard visualizing agents such as vanillin and a UV fluorescence Spectroline (254 nm) lamp. Melting points were obtained in a Fisher-Jones apparatus and are uncorrected (°C). NMR spectra were recorded in: (200 MHz)



Scheme 2.

Varian Gemini FT 200A, (300 MHz) Eclipse JEOL, (300 MHz) UNITY 300; TMS was used as an internal standard and CDCl_3 as solvent. The IR spectra were recorded in a Nicolet FT-IR Magna 750 apparatus. MS spectra were obtained in a JEOL JEM-AX505HA (IE, 70 eV) apparatus; only the molecular ions (M^+) and major peak are reported (as percentages of the base peak).

Baylis-Hillman Reaction (General Conditions) [7]. A solution of substrate aldehyde (1 mmol) and methyl acrylate (3 mmol) in 10 mL of 1,4-dioxane/water (1:1, v/v) was stirred at room temperature in the presence of 100 mol % DABCO, and the reaction progress was monitored by TLC. Upon completion, the reaction mixture was partitioned with ethyl acetate (150 mL) and water (80 mL). The organic phase was washed with brine (2 x 50 mL), dried over anhydrous Na_2SO_4 , filtered, and concentrated under reduced pressure. The crude product was purified by flash column chromatography on silica gel, using ethyl acetate and hexane as the eluting solvents to give the desired product.

Compounds 4a [8], 4b [9], 4c [8], 4d [10], and 4f [11] are known compounds, and their spectroscopic data matched those reported in the literature. 4e. Yellow oil, rf: 0.25 (hexane AcOEt, 8:2); ^1H NMR (200 MHz, CDCl_3) δ : 3.11 (s, 1H), 3.74 (s, 3H), 5.62 (s, 1H), 5.84 (dd, $J=1.2, 1.2$ Hz, 1H), 6.31 (dd, $J=0.6, 1.0$ Hz, 1H), 7.02 (dd, $J=4.8, 1.4$ Hz, 1H), 7.20-7.30 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ : 51.95, 69.88, 121.82, 125.95, 126.17, 141.62, 142.91, 166.79; IR (cm^{-1}) 1718 (C=O); 3451, (O-H); ME (IE, m/z) (%) (M) 198 (44), 85 (100).

Xanthate addition (General Procedure)

A mixture of the Baylis-Hillman adduct (1 mmol) and 0.5 M 9-BBN solution in THF (1.5 mmol) was stirred for 10 min at room temperature (until hydrogen production ceased), under a nitrogen atmosphere. The THF was then evaporated and the resulting mixture was stirred for 48 hours with the potassium

O-ethyl dithiocarbonate (2 mmol) and acetic acid (1 mmol) in dry acetonitrile (200 ml). When the reaction was completed (as determined by TLC monitoring), the acetonitrile was evaporated, poured into H_2O , and extracted with EtOAc (3x100 ml). The combined organic layers were washed with H_2O and brine, dried (anhydrous Na_2SO_4), and evaporated under vacuum. The crude product was purified by column chromatography (silica gel). Two mmol of 9-BBN was added to the pyridine substrate.

Xanthate 4a: Eluted with 7:3 hexane-ethyl acetate. Rf: 0.36; orange oil; ^1H NMR (200 MHz, CDCl_3) δ : 1.41 (t, 3H, $J=7.2$ Hz), 2.69 (bs, 1H), 3.38-3.52 (m, 3H), 3.58 (s, 3H), 4.65 (q, 2H, $J=7.2$ Hz), 5.63 (d, 1H, $J=3.8$ Hz), 7.43-7.51 (m, 1H), 7.61-7.74 (m, 2H), 8.00 (dd, 1H, $J=8.1, 1.3$ Hz); ^{13}C NMR (75 MHz, CDCl_3) δ : 13.60, 32.04, 49.78, 52.40, 69.59, 70.06, 125.11, 128.85, 129.23, 136.20, 147.56, 173.41, 214.17; IR (film) 1050, 1221, 1736, 3497 cm^{-1} ; MS (EI) m/z 360 (3) [M^+], 29 (100), 77 (83), 220 (69), 188 (66), 104 (64) 55 (57). HRMS (FAB+) calcd for $\text{C}_{14}\text{H}_{18}\text{O}_6\text{N}_1\text{S}_2$ 360.0576, found 360.0574.

4a. Minor product from the mixture with the major product- ^1H NMR (200 MHz, CDCl_3) δ : 1.41 (t, 3H, $J=7.2$ Hz), 2.69 (bs, 1H), 3.38-3.52 (m, 3H), 3.58 (s, 3H), 4.65 (q, 2H, $J=7.2$ Hz), 5.86 (d, 1H, $J=3.8$ Hz), 7.40-7.49 (m, 1H), 7.60-7.76 (m, 2H), 8.06 (dd, 1H, $J=8.1, 1.3$ Hz).

Xanthate 4b: Eluted with 7:3 hexane-ethyl acetate Rf: 0.49; yellow solid; mp. 44-46 $^\circ\text{C}$; ^1H NMR (300 MHz, CDCl_3) δ : 1.39 (t, 3H, $J=7.1$ Hz), 3.25-3.42 (m, 3H), 3.64 (s, 3H), 4.60 (q, 2H, $J=7.2$ Hz), 5.25 (s, 1H), 7.00-7.07 (m, 1H), 7.14-7.19 (m, 1H), 7.25-7.33 (m, 1H), 7.40-7.45 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ : 13.60, 34.29, 50.87, 52.11, 68.87, 70.22, 115.21, 115.50, 124.35, 127.58, 127.92, 128.10, 129.59, 129.70, 158.07, 161.33, 173.21, 213.43; IR (film) 1050, 1225, 1735, 3476 cm^{-1} ; MS (EI) m/z 333 (3) [M^+], 87 (100), 210 (97), 193 (50), 125 (49), 315 (35). HRMS (FAB+) calcd for $\text{C}_{14}\text{H}_{18}\text{O}_4\text{F}_1\text{S}_2$ 333.0631, found 333.0630.

4b. Minor product from the mixture with the major product- ^1H NMR (300 MHz, CDCl_3) δ : 1.31 (t, 3H, $J=7.1$ Hz), 3.25-3.42 (m, 3H), 3.71 (s, 3H), 4.60 (q, 2H, $J=7.2$ Hz), 5.43 (s, 1H), 7.01-7.08 (m, 1H), 7.14-7.21 (m, 1H), 7.26-7.34 (m, 1H), 7.49-7.55 (m, 1H).

Xanthate 4c: Eluted with 96:4 ethyl acetate-methanol Rf: 0.39; yellow oil; ^1H NMR (300 MHz, CDCl_3) δ : 1.41 (t, 3H, $J=7.1$ Hz), 3.24-3.50 (m, 3H), 3.62 (s, 3H), 4.63 (q, 2H, $J=7.2$ Hz), 5.02 (d, 1H, $J=4.8$ Hz), 7.34 (d, 2H, $J=5.7$ Hz), 8.52-8.54 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ : 13.66, 34.48, 51.17, 52.11, 70.43, 70.60, 121.36, 148.75, 151.83, 172.48, 213.78; IR (film) 1050, 1222, 1737; 3224 cm^{-1} ; MS (EI) m/z 316 (2) [M^+], 108 (100), 89 (53), 57 (47), 180 (17). HRMS (FAB+) calcd for $\text{C}_{13}\text{H}_{18}\text{O}_4\text{N}_1\text{S}_2$ 316.0677, found 316.0670.

4c. Minor product from the mixture with the major product- ^1H NMR (300 MHz, CDCl_3) δ : 1.33 (t, 3H, $J=7.1$ Hz), 3.12-3.45 (m, 3H), 3.67 (s, 3H), 4.55 (q, 2H, $J=7.2$ Hz), 5.19 (d, 1H, $J=4.8$ Hz), 7.40 (d, 2H, $J=5.7$ Hz), 8.52-8.54 (m, 2H).

Xanthate 4d: Eluted with 7:3 hexane-ethyl acetate Rf: 0.42; yellow solid; mp. 74-75 °C; ¹H NMR (300 MHz, CDCl₃) δ: 1.40 (t, 3H, *J* = 7.2 Hz), 2.86 (bs, 1H), 3.07-3.13 (dd, 1H, *J* = 5.4, 5.7 Hz), 3.34-3.56 (m, 2H), 3.72 (s, 3H), 4.62 (q, 2H, *J* = 7.2 Hz), 4.55 (m, 1H), 6.23 (dd, 1H, *J* = 15.8, 6.2 Hz), 6.67 (dd, 1H, *J* = 15.8, 1.2 Hz), 7.24-7.39 (m, 5H); ¹³C NMR (75 MHz, CDCl₃) δ: 13.64, 29.62, 50.82, 51.96, 70.12, 72.96, 126.65, 127.94, 128.56, 128.79, 132.30, 136.38, 172.91, 213.96; IR (film) 1049, 1222, 1734, 3455 cm⁻¹; MS (EI) *m/z* 340 (2) [M⁺], 218 (100), 133 (69), 87 (52), 131 (45) 55 (42). HRMS (FAB⁺) calcd for C₁₆H₂₀O₄S₂ 340.0803, found 340.0807.

4d. Minor product from the mixture with the major product- ¹H NMR (300 MHz, CDCl₃) δ: 1.36 (t, 3H, *J* = 7.2 Hz), 2.86 (bs, 1H), 3.07-3.13 (dd, 1H, *J* = 5.4, 5.7 Hz), 3.23-3.44 (m, 2H), 3.70 (s, 3H), 4.62 (q, 2H, *J* = 7.2 Hz), 5.18 (m, 1H), 6.25 (dd, 1H, *J* = 15.8, 6.2 Hz), 6.62 (dd, 1H, *J* = 15.8, 1.2 Hz), 7.24-7.39 (m, 5H).

Xanthate 4e: Eluted with 7:3 hexane-ethyl acetate Rf: 0.5; yellow solid; mp. 44-46 °C; ¹H NMR (200 MHz, CDCl₃) δ: 1.40 (t, 3H, *J* = 7.1 Hz), 2.63 (bs, 1H), 3.24-3.44 (m, 3H), 3.69 (s, 3H), 4.63 (q, 2H, *J* = 7.2 Hz), 5.04 (d, 1H, *J* = 5.4 Hz), 7.07 (dd, 1H, *J* = 4.8, 1.4 Hz), 7.24-7.36 (m, 2H); ¹³C NMR (50 MHz, CDCl₃) δ: 13.69, 34.40, 51.18, 52.19, 70.26, 70.95, 121.97, 125.51, 126.58, 142.45, 173.42, 213.78; IR (film) 1048, 1221, 1733, 3471 cm⁻¹; MS (EI) *m/z* 319 (4) [MH⁻], 198 (100), 181 (92), 85 (70), 302 (60). HRMS (FAB⁺) calcd for C₁₂H₁₆O₄S₃ 320.0211, found 320.0208.

4e. Minor product from the mixture with the major product- ¹H NMR (200 MHz, CDCl₃) δ: 1.39 (t, 3H, *J* = 7.1 Hz), 2.63 (bs, 1H), 3.20-3.48 (m, 3H), 3.69 (s, 3H), 4.58 (q, 2H, *J* = 7.2 Hz), 5.20 (d, 1H, *J* = 5.4 Hz), 7.07 (dd, 1H, *J* = 4.8, 1.4 Hz), 7.24-7.36 (m, 2H).

Xanthate 4f: Eluted with 7:3 hexane-ethyl acetate Rf: 0.43; yellow oil; ¹H NMR (300 MHz, CDCl₃) δ: 1.41 (t, 3H, *J* = 7.2 Hz), 2.17 (s, 1H), 3.18-3.50 (m, 3H), 3.72 (s, 3H), 4.64 (q, 2H, *J* = 7.2 Hz), 4.94 (d, 1H, *J* = 5.4 Hz), 6.39-6.40 (m, 1H), 7.41-7.43 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ: 13.73, 34.37, 50.54, 52.22, 67.52, 70.31, 108.42, 126.08, 139.74, 143.68, 173.42, 213.77; IR (film) 1051, 1220, 1738, 3454 cm⁻¹; MS (EI) *m/z* 318 (2) [M⁺], 182 (100), 87 (29), 97 (15).

4f. Minor product from the mixture with the major product- ¹H NMR (300 MHz, CDCl₃) δ: 1.38 (t, 3H, *J* = 7.2 Hz), 2.17 (s, 1H), 3.18-3.50 (m, 3H), 3.70 (s, 3H), 4.60 (q, 2H, *J* = 7.2 Hz), 5.53 (d, 1H, *J* = 5.4 Hz), 6.39-6.40 (m, 1H), 7.38-7.40 (m, 2H).

6e (Alcohol protection)

A mixture of the xanthate (1 mmol), imidazol (2 mmol), and TBSCl (1.1 mmol) in DMF (2ml/g de MP), was stirred at room temperature under a nitrogen atmosphere for 5 days. When the reaction was completed (monitored by TLC), the mixture was

poured into H₂O (10 ml), and extracted with EtOAc (3 x 5 ml). The combined organic layers were washed with H₂O, brine, dried (anhyd Na₂SO₄), and evaporated. The crude product was purified by column chromatography (silica gel). The colorless product was obtained by slow precipitation from methanol/hexane. Eluted with 8:2 hexane-ethyl acetate Rf: 0.55; white solid; mp. 53-54 °C; ¹H NMR (200 MHz, CDCl₃) δ: 0.24 (s, 3H), -0.01 (s, 3H), 0.80 (s, 9H), 1.33 (t, 3H, *J* = 7.2 Hz), 2.91-3.21 (m, 3H), 3.69 (s, 3H), 4.55 (q, 2H, *J* = 7.1 Hz), 4.95 (d, 1H, *J* = 7.8 Hz), 7.03-7.06 (dd, 1H, *J* = 5.1, 1.3 Hz), 7.13-7.16 (m, 1H), 7.27-7.31 (m, 1H); ¹³C NMR (50 MHz, CDCl₃) δ = -5.40, -4.91, 13.72, 17.98, 25.56, 33.86, 51.88, 53.58, 70.01, 72.35, 122.29, 126.01, 126.15, 142.64, 172.83, 213.59; IR (film) 839, 1051, 1085, 1221, 1739, C=O cm⁻¹; MS (EI) *m/z* 434 (3) [MH⁺], 255(100).

Acknowledgements

Financial support from the DGAPA-UNAM (project PAPIIT-IN213407) is gratefully acknowledged and Dr. Joseph M. Muchowski for many helpful discussions. Also we thank H. Rios, R. Patiño, J. Pérez, L. Velasco, N. Zavala, E. Hernández, M. I. Chávez and A. Peña for technical support and R. A. Toscano and S. Hernández for X-ray crystallography.

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