

# A Green Synthesis of 3, 4-Dihydropyrimidin-2(1H)-ones via One-Pot Multi-Component Reaction by Using Cuttlebone as a Natural Catalyst under Solvent-Free Conditions

Nooshin Firoozeh, Soodabeh Rezazadeh\* and Cobra Izanloo

Department of Chemistry, Faculty of Sciences, Bojnourd Branch, Islamic Azad University, Bojnourd 9417697796, Iran.  
soodabehrezazadeh@yahoo.com

Received February 14<sup>th</sup>, 2017; Accepted May 12<sup>th</sup>, 2017.

**Abstract.** In this paper, an efficient, green and eco-friendly method for synthesis of 3,4-dihydropyrimidin-2(1H)-ones via one-pot multi-component reaction of aldehydes, urea and ethyl acetoacetate is shown at the presence of cuttlebone as an effective, reusable and natural heterogeneous catalyst. The advantages of this method are short reaction times, high yields, simple work-up, and reusable natural catalysts. The structure of cuttlebone was characterized by FT-IR spectrum, scanning electron microscopy (SEM) and energy dispersive spectrometer (SEM-EDS).

**Key words:** Cuttlebone; 3, 4-dihydropyrimidin-2(1H)-ones; aldehydes; one-pot reaction; green.

**Resumen:** En este documento, se muestra un método eficiente, agradable para el medio ambiente y ecológico para la síntesis de 3,4-dihydropyrimidin-2 (1H)-onas, mediante la reacción de multicomponentes de aldehídos, urea y acetato de etilo en presencia de "cuttlebone" como un catalizador heterogéneo natural, eficaz y reutilizable. Unas de las ventajas de este método son: tiempos cortos de reacción, alta eficiencia, procedimiento fácil y el empleo de un catalizador natural y reutilizable. La estructura del "cuttlebone" fue caracterizada por FT-IR, microscopía electrónica de barrido (SEM) y espectrometría de dispersión de energía (SEM-EDS).

**Palabras clave:** Cuttlebone; 3, 4-dihidropirimidin-2(1H)-onas; aldehídos; reacción en un solo recipiente; química verde.

## Introduction

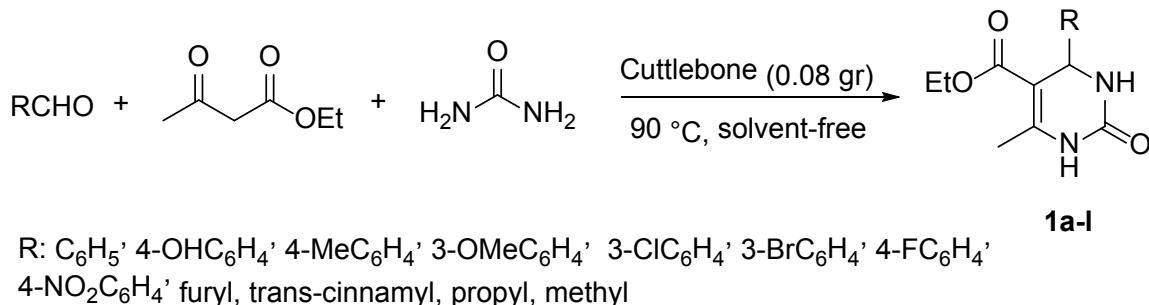
3, 4-Dihydropyrimidin-2(1H)-ones are an important class of heterocyclic compounds in the area of natural and synthetic organic chemistry. They possess a variety curative and pharmacological applications involving antiviral, antitumor, antibacterial and anti-inflammatory activities [1-3]. Moreover, these compounds have appeared as potential calcium channel blockers [4], antihypertensive [5],  $\alpha$ 1a-adrenergic antagonists [6] and neuropeptide antagonists [7].

Multi-component reactions (MCRs) are one-pot reactions in which three or more starting materials reacted to form a product. MCRs display high atom economy and high selectivity [8-10]. The advantages of these reactions involve: synthetic productivity, simplicity of procedure, decreasing number of isolation and purification steps, save on energy, time and costs and also reduction of waste [11-14]. The synthesis of 3, 4-dihydropyrimidin-2(1H)-ones (DHPMs) which was initially reported by Pietro Biginelli is one of the excellent MCRs that includes a three-component one-pot reaction of an aldehyde, ethyl acetoacetate and urea in the presence of strong acid [15]. The disadvantages of classical Biginelli reaction include hard reaction conditions, long reaction times and low yields [16-18]. In recent years, new methods for preparation of DHPMs have been developed to improve and modify this reaction using microwave irradiation [19], ultrasound irradiation [20], ionic liquids

[21], Lewis and Bronsted acid such as lanthanide triflate [22],  $H_3BO_3$  [23],  $VCl_3$  [24],  $Sr(OTf)_2$  [25],  $Bi_2(SO_4)_3$  [26], Indium(III) halides [27],  $KAl(SO_4)_2 \cdot 12H_2O$  supported on silica [28], silicasulfuric acid [23,29],  $Mn(OAc)_3 \cdot 2H_2O$  [30],  $Y(NO_3)_3 \cdot 6H_2O$  [31], zeolite [32],  $TaBr_5$  [33],  $Ce(NO_3)_3 \cdot 6H_2O$  [34], glutamic acid [35],  $H_7[PMo_8V_4O_{40}]$  [36], lemon juice [37] and so on. However, some of the methods are presented to have one or more drawbacks such the use of expensive reagents, harmful to the environment, strong acidic conditions, long reaction times and low yields. Due to the importance of DHPMs, the introduction of mild and green methods with higher yields are needed.

Green chemistry is the model of chemical products and processes that reduce or eliminate the use or generation of hazardous substance. Nowadays the development of eco-friendly and economical processes is a challenge for chemists. Replacement of toxic, polluting and non-recyclable catalysts with eco-friendly reusable solid, heterogeneous catalysts are an area of current interest [38].

According to the above points, in this research, we want to report a convenient and green method for the synthesis of 3,4-dihydropyrimidin-2(1H)-ones via the one-pot multi-component reaction of aldehydes with ethyl acetoacetate and urea at the presence of cuttlebone as an eco-friendly, recyclable and heterogeneous natural catalyst under solvent-free conditions (Scheme 1).



**Scheme 1.** The synthesis of 3, 4-dihydropyrimidin-2(1*H*)-ones using cuttlebone as a green catalyst.

## Results and Discussion

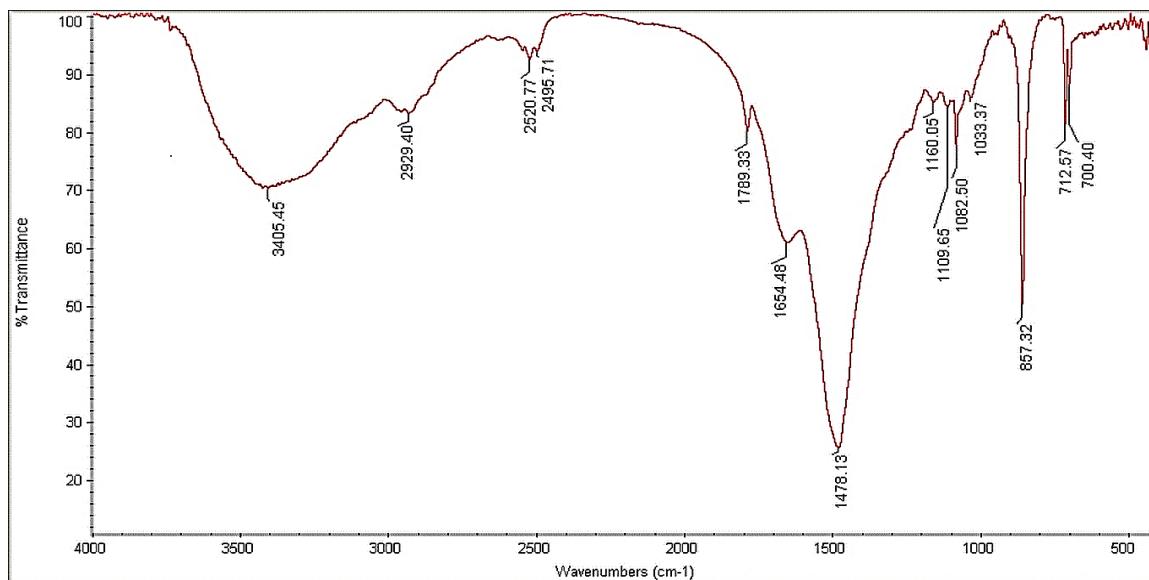
In this paper, we describe a simple and efficient protocol for the synthesis of 3,4-dihydropyrimidin-2(1*H*)-ones *via* one-pot multi-component condensation of aldehydes, ethyl acetoacetate and urea using cuttlebone as a novel, reusable and eco-friendly heterogeneous natural catalyst.

### Characterization of cuttlebone

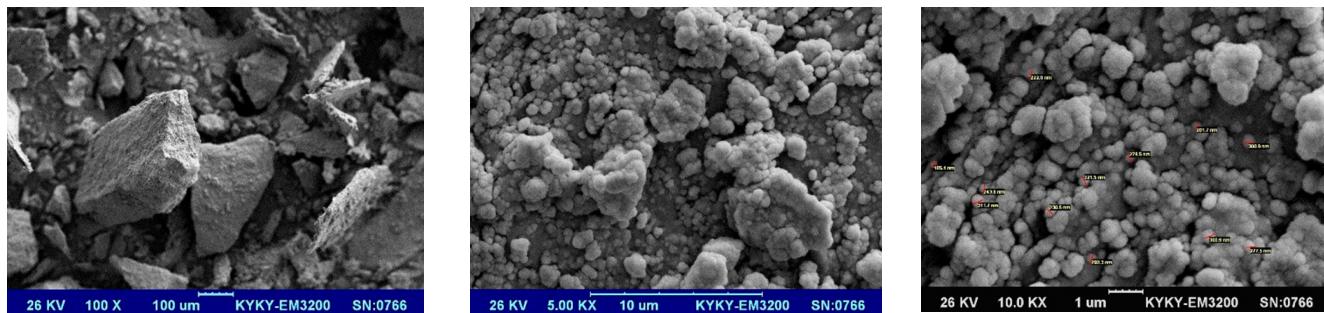
Cuttlebone is the calcic internal shell of a cuttlefish. The cuttlebone is a class of ultra-lightweight cellular natural material that possess unique chemical, mechanical and structural properties such as high rigidity, high porosity and high penetrance. The dorsal shield and the lamellar matrix are two main components of cuttlebone [39]. The dorsal shield has a hard and stiff structure that provides a rigid stratum for protection of the structure and the development of the lamellar matrix of cuttlebone [40,41]. The lamellar matrix of cuttlebone has the highest

degree of porosity (up to 90%), but also can tolerate high hydrostatic pressure. The lamellar matrix consists primarily of aragonite (a crystallized form of calcium carbonate,  $\text{CaCO}_3$ ), covered in a layer of organic material [40-42] composed of  $\beta$ -chitin [43]. Cuttlebone that is also known as a cuttlefish bone (*Sepia esculenta*) [44-47] is commonly found on saltwater beaches like the Persian Gulf in Iran.

The FT-IR spectrum of powdered cuttlebone is shown in Fig. 1. The absorption bands of planar  $\text{CO}_3^{2-}$  ion (aragonite) are observed at 1082, 857, 1478 and 700  $\text{cm}^{-1}$ . Also, the board bands at about 3500-3300  $\text{cm}^{-1}$  are attributed to the OH and NH stretching vibrations of  $\beta$ -chitin segment of cuttlebone, respectively. In fact, the hydrogen bonding network of  $\beta$ -chitin of cuttlebone cause to broadening absorption bands of OH and NH groups. The absorption bands of symmetric stretching of CH,  $\text{CH}_3$  and asymmetric stretching vibrations of  $\text{CH}_2$ , are appeared from 2929 to 2855  $\text{cm}^{-1}$ . In addition, the stretching vibration of C=O of amide at 1654  $\text{cm}^{-1}$  and the stretching vibrations of asymmetric bridge oxygen and C–O at 1160 to 1030  $\text{cm}^{-1}$ , are observed [48-50].



**Fig. 1.** The FT-IR spectrum of powdered cuttlebone.



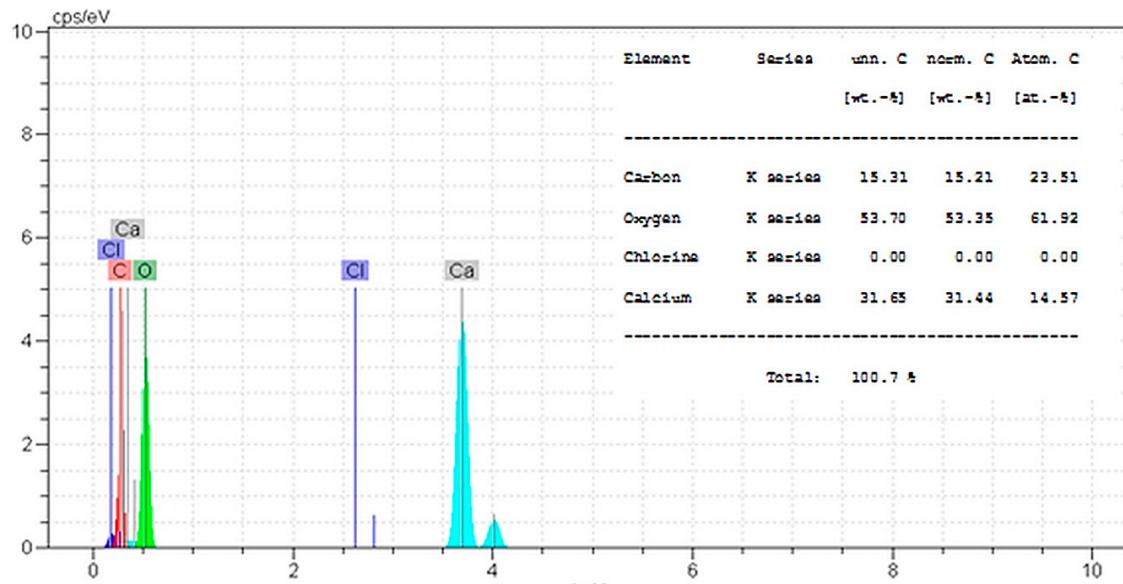
**Fig. 2.** The SEM images of powdered cuttlebone.

Scanning electronic microscopy (SEM) of powdered cuttlebone shows an amorphous morphology with a 185-321 nm particle size (Fig. 2).

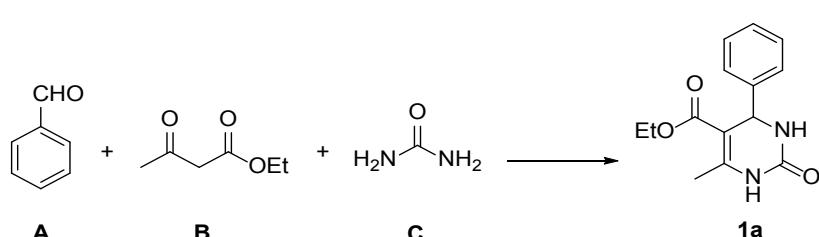
Fig. 3 shows the energy dispersive spectrum (EDS) of powdered cuttlebone. According to this spectrum, the cuttlebone structure consists of Ca, C and O elements.

In order to find the best reaction conditions, the one-pot three-component reaction of benzaldehyde, ethyl acetoacetate and urea was chosen as a model reaction (Scheme 2).

At first the model reaction was studied at the presence of 0.1 gr of some natural catalysts such as cuttlebone, calcined eggshell (CES) and natural zeolite under solvent-free conditions at 100 °C (Table 1, entries 1-3). As Table 1 (entry 1) shows the use of cuttlebone is more efficient for the model reaction. Then, the different amounts of cuttlebone was tested on the model reaction (Table 1, entries 1 and 4-5). According to the results, increasing the amount of catalyst has no effect on yield of product, while a decrease of the amount of the catalyst cause



**Fig. 3.** The EDS spectrum of powdered cuttlebone.



**Scheme 2.** The one-pot three-component reaction of benzaldehyde, ethyl acetoacetate and urea under different conditions.

**Table 1.** The effect of varying conditions on the model reaction.

Entry <sup>a</sup>	Catalyst	Catalyst (gr)	Molar ratio		Solvent	Temp. (°C)	Time (min)	Yield (%)
			A: B: C					
1	Cuttlebone	0.1	1:1:1		s.f <sup>b</sup>	100	10	92
2	CES	0.1	1:1:1		s.f <sup>b</sup>	100	30	90
3	Natural Zeolite	0.1	1:1:1		s.f <sup>b</sup>	100	40	85
4	Cuttlebone	0.08	1:1:1		s.f <sup>b</sup>	100	10	92
5	Cuttlebone	0.06	1:1:1		s.f <sup>b</sup>	100	20	90
6	Cuttlebone	0.08	1:1:1		s.f <sup>b</sup>	90	10	92
7	Cuttlebone	0.08	1:1:1		s.f <sup>b</sup>	80	30	85
8	Cuttlebone	0.08	1:1:1.2		s.f <sup>b</sup>	90	10	92
9	Cuttlebone	0.08	1:1:1		MeOH	reflux	3 h	-
10	Cuttlebone	0.08	1:1:1		CH <sub>3</sub> CN	reflux	3 h	80
11	Cuttlebone	0.08	1:1:1		H <sub>2</sub> O	reflux	3 h	-

<sup>a</sup> Isolated yield; <sup>b</sup> Solvent-free.

of decreasing desired product (**1a**). After that, the model reaction was carried out under different temperature at the presence of 0.08 gr of catalyst (Table 1, entries 4, 6-7). The results showed that the reducing the temperature to 90 °C had no effect on yield and reaction time. Applying 1: 1: 1.2 molar ratio of benzaldehyde: ethyl acetoacetate: urea at the presence of 0.08 gr of catalyst under solvent-free conditions had no effect on yield and reaction time (Table 1, entries 6 and 8). Finally, to investigate the effect of solvent, the model reaction was performed in presence and absence of solvent (Table 1, entries 6 and 9-11). The best results were obtained with 0.08 gr of cuttlebone under solvent-free conditions at 90 °C (Table 1, entry 6).

To indicate the scope and the generality of the method, ethyl acetoacetate was reacted with aromatic, heteroaromatic and aliphatic aldehydes with a wide range of functional groups and urea under an optimized reaction conditions to giving 3, 4-dihydropyrimidin-2(1*H*)-ones with good to high yields (75-95%). The results are presented in Table 2.

Aromatic aldehydes with unsubstituted/substituted electron-donating and electron-withdrawing groups such as -OH, -CH<sub>3</sub>, -OCH<sub>3</sub>, -Cl, -Br, -F, -NO<sub>2</sub> gave excellent yields of corresponding products (Table 3, entries 1-8). Furthermore,  $\alpha$ ,  $\beta$ -unsaturated aldehydes and heteroaromatic aldehydes also afforded good yields (Table 3, entries 9 and 10). Interestingly, aliphatic aldehydes also gave desired products in good yields. (Table 3, entries 11 and 12).

To check the recyclability of catalyst, after completion of the reaction of benzaldehyde, ethyl acetoacetate with urea, the catalyst was separated from the reaction mixture, washed with ethanol and distilled water several times to remove the organic products, and dried. The heterogeneous catalyst was used for 3 successive times in the new experiments obtaining good yields (Fig. 4).

## Conclusion

We have developed a novel and green protocol for the synthesis of 3, 4-dihydropyrimidin-2(1*H*)-ones via one-pot multi-component reaction. Generality, efficiency, simple work-up procedure and purification, recyclability of the catalyst, short reaction times, high yields, low cost and finally conformity with the green chemistry principles are the advantages of this protocol.

## Experimental

The chemicals were purchased from Merck. Melting points of products were determined with an Electrothermal Type 9100 melting point apparatus. The FT-IR spectra were recorded on an Avatar 370 FT-IR Thermo Nicolet spectrometer. The NMR spectra were determined on a Bruker Avance 300 MHz instrument in DMSO-*d*<sub>6</sub>. Elemental analyses were performed using an Elementar, Vario EL III and Thermofinnigan Flash EA 1112 Series instrument. All the reactions were monitored by thin layer chromatography (TLC); the spots were visualized with UV light. Scanning electron microscopy (SEM) and energy dispersive spectrometry (SEM-EDS) were performed on a KYKY-EM3200 operated at a 30 KV accelerating. All of the products were known compound and characterized by the FT-IR spectroscopy. The structure of products was further confirmed by <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopy. Cuttlebone, that is also known as cuttlefish bone (*Sepia esculenta*) [44-47] is commonly found on saltwater beaches like the Persian Gulf in Iran.

### Pre-preparation of cuttlebone

In order to the use of cuttlebone in reaction, at first the catalyst was powdered, then washed with distilled water to remove

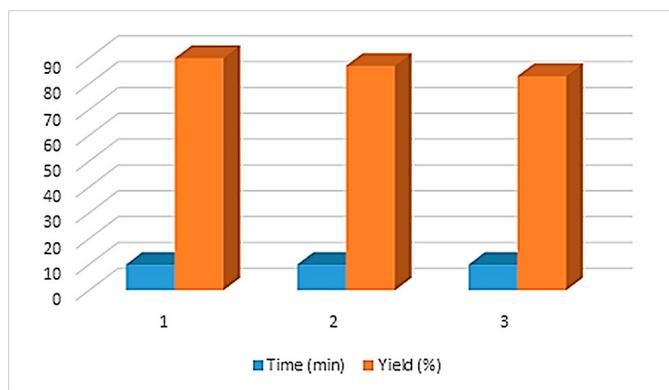
**Table 2.** Synthesis of 3, 4-dihydropyrimidin-2(1*H*)-ones (**1a-1l**) catalyzed by cuttlebone under solvent-free conditions.

Entry	Aldehyde	Product	Time (min)	Yield <sup>a</sup> (%)
1			10	92
2			30	90
3			60	95
4			60	89
5			15	94
6			15	95

**Table 2.** Synthesis of 3, 4-dihydropyrimidin-2(1*H*)-ones (**1a-1l**) catalyzed by cuttlebone under solvent-free conditions.

Entry	Aldehyde	Product	Time (min)	Yield <sup>a</sup> (%)
7			15	92
8			60	85
9			40	87
10			60	90
11			60	86
12			60	75

<sup>a</sup> Isolated yield



**Fig. 4.** Synthesis of 5-ethoxycarbonyl-4-phenyl-6-methyl-3, 4-dihydropyrimidin-2(1*H*)-one (**1a**) in the presence of reused catalyst.

pollution on the surface of cuttlebone and finally dried at 100 °C for 2 h [48].

**Typical procedure for the preparation of 5-(ethoxycarbonyl)-6-methyl-4-phenyl-3, 4-dihydropyrimidin-2(1*H*)-one (**1a**).**

A mixture of benzaldehyde (1 mmol, 0.106 gr), ethyl acetoacetate (1 mmol, 0.147 gr), urea (1 mmol, 0.060 gr) and cuttlebone (0.08 gr) was heated at 90 °C in an oil bath for 10 min. After completion of the reaction as monitored by TLC, the reaction mixture was cooled to room temperature, diluted with ethanol (10 mL) and filtered to recover the catalyst. The organic layer was separated and concentrated under reduced pressure. The crude product was recrystallized from ethanol to afford 0.241 gr **1a** (92%).

**5-(Ethoxycarbonyl)-6-methyl-4-phenyl-3,4-dihydropyrimidin-2(1*H*)-one (**1a**).** White solid; mp 206–208 °C (Lit. 206–208 °C) [51]; FT-IR (KBr)  $\nu_{max}$  3244, 3116, 2978, 2937, 1725, 1702, 1648, 1596, 1465, 1421, 1313, 1221, 1090, 1026, 783, 757, 699, 661, 516 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.22 (1H, s, NH), 7.77 (1H, s, NH), 7.25–7.36 (5H, m, C<sub>6</sub>H<sub>5</sub>), 5.16 (1H, d, *J* = 3.0 Hz, CH), 4.00 (2H, q, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.27 (3H, s, CH<sub>3</sub>), 1.11 (3H, t, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ 165.7, 152.6, 148.8, 145.3, 128.8, 127.7, 126.7, 99.7, 59.6, 54.4, 18.7, 14.5. *Anal.* C 64.57%, H 6.12%, N 10.71%, calcd for C<sub>14</sub>H<sub>16</sub>N<sub>2</sub>O<sub>3</sub>, C 64.60%, H 6.20%, N 10.76%.

**5-(Ethoxycarbonyl)-4-(4-hydroxyphenyl)-3,4-dihydropyrimidin-2(1*H*)-one (**1b**).** White solid; mp 228–230 °C (Lit. 228–230 °C) [52]; FT-IR (KBr)  $\nu_{max}$  3513, 3362, 3278, 3239, 3118, 2982, 2814, 1685, 1649, 1596, 1516, 1461, 1383, 1317, 1229, 1171, 1091, 825, 753, 651, 544, 507 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.77 (2H, s, NH and OH), 9.32 (1H, s, NH), 7.09 (2H, d, *J* = 8.9 Hz, C<sub>6</sub>H<sub>5</sub>), 6.74 (2H, d, *J* = 8.9 Hz, C<sub>6</sub>H<sub>5</sub>), 5.12 (1H, s, CH), 3.97 (2H, q, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.27 (3H, s, CH<sub>3</sub>), 1.08 (3H, t, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); *Anal.* C

60.79%, H 5.66%, N 10.09%, calcd for C<sub>14</sub>H<sub>16</sub>N<sub>2</sub>O<sub>4</sub>, C 60.86%, H 5.84%, N 10.14%.

**5-(Ethoxycarbonyl)-6-methyl-4-(4-methylphenyl)-3,4-dihydropyrimidin-2(1*H*)-one (**1c**).** White solid; mp 209–211 °C (Lit. 209–211 °C) [53]; FT-IR (KBr)  $\nu_{max}$  3245, 3115, 2980, 2953, 1728, 1704, 1649, 1461, 1422, 1328, 1287, 1222, 1089, 785, 478 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.17 (1H, s, NH), 7.70 (1H, s, NH), 7.13 (4H, s, C<sub>6</sub>H<sub>4</sub>), 5.11 (1H, d, *J* = 3.0 Hz, CH), 3.99 (2H, q, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.27 (3H, s, CH<sub>3</sub>), 2.25 (3H, s, CH<sub>3</sub>), 1.11 (3H, t, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ 165.8, 152.6, 148.6, 142.4, 136.8, 129.3, 126.6, 99.8, 59.6, 54.1, 21.1, 18.2, 14.6. *Anal.* C 65.70%, H 6.58%, N 10.20%, calcd for C<sub>15</sub>H<sub>18</sub>N<sub>2</sub>O<sub>3</sub>, C 65.68%, H 6.61%, N 10.21%.

**5-(Ethoxycarbonyl)-6-methyl-4-(3-methoxyphenyl)-3,4-dihydropyrimidin-2(1*H*)-one (**1d**).** White solid; mp 200–203 °C (Lit. 200–202 °C) [54]; FT-IR (KBr)  $\nu_{max}$  3449, 3268, 3245, 3116, 3014, 2961, 2937, 1704, 1651, 1599, 1493, 1330, 1286, 1227, 1094, 1039, 790, 744, 453 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.20 (1H, s, NH), 7.74 (1H, s, NH), 7.26 (1H, t, *J* = 6.0 Hz, C<sub>6</sub>H<sub>4</sub>), 6.85–6.79 (3H, m, C<sub>6</sub>H<sub>4</sub>), 5.14 (1H, d, *J* = 3.0 Hz, CH), 4.01 (2H, q, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 3.73 (3H, s, OCH<sub>3</sub>), 2.25 (3H, s, CH<sub>3</sub>), 1.12 (3H, t, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); *Anal.* C 64.00%, H 6.21%, N 9.47%, calcd for C<sub>15</sub>H<sub>18</sub>N<sub>2</sub>O<sub>4</sub>, C 64.06%, H 6.25%, N 9.65%.

**5-(Ethoxycarbonyl)-4-(3-chlorophenyl)-6-methyl-3,4-dihydropyrimidin-2(1*H*)-one (**1e**).** White solid; mp 190–192 °C (Lit. 190–192 °C) [52]; FT-IR (KBr)  $\nu_{max}$  3438, 3301, 3223, 3120, 2980, 2928, 1707, 1662, 1612, 1588, 1528, 1475, 1376, 1286, 1227, 1122, 1090, 1022, 870, 783, 691, 605, 446 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.29 (1H, s, NH), 7.82 (1H, s, NH), 7.41–7.20 (4H, m, C<sub>6</sub>H<sub>4</sub>), 5.16 (1H, s, CH), 4.01 (2H, q, *J* = 3.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.27 (3H, s, CH<sub>3</sub>), 1.11 (3H, t, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); *Anal.* C 56.97%, H 5.12%, N 9.39%, calcd for C<sub>14</sub>H<sub>15</sub>ClN<sub>2</sub>O<sub>3</sub>, C 57.05%, H 5.13%, N 9.50%.

**5-(Ethoxycarbonyl)-4-(3-bromophenyl)-6-methyl-3,4-dihydropyrimidin-2(1*H*)-one (**1f**).** White solid; mp 196–199 °C (Lit. 196–198 °C) [55]; FT-IR (KBr)  $\nu_{max}$  3436, 3299, 3239, 3120, 2976, 2937, 1705, 1658, 1612, 1589, 1527, 1473, 1377, 1285, 1226, 1090, 782, 693, 603, 456 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.34 (1H, s, NH), 7.86 (1H, s, NH), 7.35–7.40 (4H, m, C<sub>6</sub>H<sub>4</sub>), 5.20 (1H, d, *J* = 3.0 Hz, CH), 4.07 (2H, q, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.32 (3H, s, CH<sub>3</sub>), 1.17 (3H, t, *J* = 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ 158.0, 149.4, 146.1, 131.2, 130.7, 130.2, 129.6, 129.1, 99.1, 59.0, 54.0, 18.3, 14.5. *Anal.* C 49.51%, H 4.32%, N 8.09%, calcd for C<sub>14</sub>H<sub>15</sub>BrN<sub>2</sub>O<sub>3</sub>, C 49.58%, H 4.46%, N 8.26%.

**5-(Ethoxycarbonyl)-4-(4-fluorophenyl)-6-methyl-3,4-dihydropyrimidin-2(1*H*)-one (**1g**).** White solid; mp 176–178 °C (Lit. 176–178 °C) [52]; FT-IR (KBr)  $\nu_{max}$  3445, 3337, 3247, 3121, 2980, 2953, 2932, 1727, 1702, 1648, 1600, 1508, 1463,

1290, 1221, 1161, 1089, 840, 793, 606, 592, 508, 459 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.28 (1H, s, NH), 7.79 (1H, s, NH), 7.28 (2H, t, *J*= 9.0 Hz, C<sub>6</sub>H<sub>4</sub>), 7.15 (2H, t, *J*= 9.0 Hz, C<sub>6</sub>H<sub>4</sub>), 5.18 (1H, s, CH), 3.99 (2H, q, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.28 (3H, s, CH<sub>3</sub>), 1.09 (3H, t, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); *Anal.* C 60.23%, H 5.12%, N 10.11%, calcd for C<sub>14</sub>H<sub>15</sub>FN<sub>2</sub>O<sub>3</sub>, C 60.43%, H 5.43%, N 10.07%.

**5-(Ethoxycarbonyl)-6-methyl-4-(4-nitrophenyl)-3,4-dihydropyrimidin-2(1*H*)-one (1h).** Pall yellow solid; mp 174-178 °C (Lit. 175-177.5 °C) [56a]; FT-IR (KBr) *v*<sub>max</sub> 3485, 3314, 3108, 2978, 2937, 1732, 1711, 1677, 1601, 1568, 1527, 1371, 1351, 1298, 1244, 1177, 1106, 1014, 856, 699, 590 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.41 (1H, s, NH), 8.14 (2H, d, *J*= 9.0 Hz, C<sub>6</sub>H<sub>4</sub>), 7.50 (1H, s, NH), 7.42 (2H, d, *J*= 9.0 Hz, C<sub>6</sub>H<sub>4</sub>), 5.14 (1H, s, CH), 4.09 (2H, q, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.20 (3H, s, CH<sub>3</sub>), 1.19 (3H, t, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); *Anal.* C 54.98%, H 4.99%, N 13.63%, calcd for C<sub>14</sub>H<sub>15</sub>N<sub>3</sub>O<sub>5</sub>, C 55.08%, H 4.95%, N 13.76%.

**5-(Ethoxycarbonyl)-4-(furan-2-yl)-6-methyl-3,4-dihydropyrimidin-2(1*H*)-one (1i).** White solid; mp 194-196 °C (Lit. 196 °C) [56b]; FT-IR (KBr) *v*<sub>max</sub> 3335, 3251, 3120, 2965, 2928, 2851, 1703, 1650, 1457, 1380, 1236, 1098, 798, 465 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.17 (1H, s, NH), 7.63 (1H, d, *J*= 6.0 Hz, furyl), 7.49 (1H, s, NH), 6.28 (1H, t, *J*= 3.0 Hz, furyl), 6.02 (1H, d, *J*= 3.0 Hz, furyl), 5.13 (1H, s, CH), 3.96 (2H, q, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 2.16 (3H, s, CH<sub>3</sub>), 1.07 (3H, t, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>). *Anal.* C 57.32%, H 5.57%, N 11.21%, calcd for C<sub>12</sub>H<sub>14</sub>N<sub>2</sub>O<sub>4</sub>, C 57.59%, H 5.64%, N 11.19%.

**5-(Ethoxycarbonyl)-6-methyl-4-styryl-3,4-dihydropyrimidin-2(1*H*)-one (1g).** White solid; mp 238-241 °C (Lit. 240-242 °C) [56c]; FT-IR (KBr) *v*<sub>max</sub> 3244, 3112, 3035, 2977, 2932, 1722, 1702, 1652, 1463, 1424, 1387, 1367, 1288, 1227, 1095, 967, 779, 756, 692, 488 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.21 (1H, s, NH), 7.61 (1H, s, NH), 7.28-7.47 (5H, m, C<sub>6</sub>H<sub>5</sub>), 6.43 (1H, d, *J*= 15.0 Hz, CH vinyl), 6.25 (1H, dd, *J*<sub>1</sub>= 18.0 Hz, *J*<sub>2</sub>= 6.0 Hz, CH vinyl), 4.79 (1H, dd, *J*<sub>1</sub>= 6.0 Hz, *J*<sub>2</sub>= 3.0 Hz, CH), 4.08-4.20 (2H, m, OCH<sub>2</sub>CH<sub>3</sub>), 2.26 (3H, s, CH<sub>3</sub>), 1.26 (3H, t, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ 165.6, 153.0, 149.0, 136.6, 130.4, 129.1, 128.5, 128.0, 126.8, 98.2, 59.6, 52.3, 18.2, 14.7. *Anal.* C 66.97%, H 6.39%, N 9.72%, calcd for C<sub>16</sub>H<sub>18</sub>N<sub>2</sub>O<sub>3</sub>, C 67.12%, H 6.34%, N 9.78%.

**5-Ethoxycarbonyl-6-methyl-4-*n*-propyl-3,4-dihydropyrimidin-2(1*H*)-one (1k)** White solid; mp 177-180 °C (Lit. 178-180 °C) [56d]; FT-IR (KBr) *v*<sub>max</sub> 3362, 3251, 3121, 2986, 2958, 2937, 1721, 1698, 1674, 1645, 1482, 1464, 1369, 1332, 1286, 1235, 1119, 1090, 1016, 787, 663, 599 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 8.95 (1H, s, NH), 7.34 (1H, s, NH), 4.12-4.03 (3H, m, OCH<sub>2</sub>CH<sub>3</sub> and CH), 2.17 (3H, s, CH<sub>3</sub>), 1.45-1.24 (4H, m, CH<sub>2</sub>-CH<sub>2</sub>), 1.20 (3H, t, *J*= 6.0 Hz, CH<sub>3</sub>), 0.86 (3H, t, *J*= 6.0 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ 165.9, 153.2, 148.7, 99.8, 59.5, 50.2, 18.1, 17.4, 14.6, 14.2. *Anal.* C

58.32%, H 8.10%, N 12.25%, calcd for C<sub>11</sub>H<sub>18</sub>N<sub>2</sub>O<sub>3</sub>, C 58.39%, H 8.02%, N 12.38%.

**5-Ethoxycarbonyl-4,6-dimethyl-3,4-dihydropyrimidin-2(1*H*)-one (1l).** White solid; mp 175-178 °C (Lit. 178 °C) [56e]; FT-IR (KBr) *v*<sub>max</sub> 3438, 3345, 3260, 2929, 2804, 2639, 1682, 1625, 1601, 1465, 1153, 1053, 1002, 788, 718, 559 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 9.12 (1H, s, NH), 7.29 (1H, s, NH), 4.19-4.05 (3H, m, OCH<sub>2</sub>CH<sub>3</sub> and CH), 2.19 (3H, s, CH<sub>3</sub>), 1.25-1.17 (6H, m, OCH<sub>2</sub>CH<sub>3</sub> and CH<sub>3</sub>CH); *Anal.* C 54.19%, H 7.17%, N 14.05%, calcd for C<sub>9</sub>H<sub>14</sub>N<sub>2</sub>O<sub>3</sub>, C 54.53%, H 7.12%, N 14.13%.

## Acknowledgments

The authors gratefully acknowledge the partial support of this study by the Islamic Azad University of Bojnourd.

## References

1. Kappe, C. O. *Tetrahedron* **1993**, *49*, 6937.
2. Kappe, C. O. *Acc. Chem. Res.* **2000**, *33*, 879.
3. Kappe, C. O. *Eur. J. Med. Chem.* **2000**, *35*, 1043.
4. Atwal, K. S.; Swanson, B. N.; Unger, S. E.; Floyed, D. M.; Moreland, S.; Hedberg, A.; O'Reilly, A. *J. Med. Chem.* **1991**, *34*, 806.
5. Adib, M.; Sheikhi, E.; Kavoosi, A.; Bijanzadeh, H.R. *Tetrahedron* **2010**, *66*, 9263.
6. Pourjavadi, A.; Salimi, H.; Barzegar, Sh.; Eftekhari, B. *Acta Chim. Slov.* **2007**, *54*, 140.
7. Atwal, K. S.; Rovnyak, G. C.; O'Reilly, B. C. Schwartz, J. *J. Org. Chem.* **1989**, *54*, 5898.
8. Tao, L.; Zhu, C.; Wei, Y.; Zhao, Y. *Adv. Polym. Sci.* **2015**, *269*, 43.
9. Elnagdi, M. H; Moustafa, M. S.; Al-Mousawi, S. M.; Mekheimer, R. A.; Sadek, K. U. *Mol. Divers.* **2015**, *19*, 625.
10. Singh, M. S.; Chowdhury, S. *RSC Adv.* **2012**, *2*, 4547.
11. Zhu, J.; Bienayme, H. Eds.; *Multicomponent Reactions*, Wiley-VCH: Weinheim, Germany, 2005.
12. Dabiri, M.; Noroozi Tisseh, Z.; Bahramnejad, M.; Bazgir, A. *Ultrason. Sonochem.* **2011**, *18*, 1153.
13. Hasaninejad, A.; Zare, A.; Shekouhy, M. *Tetrahedron* **2011**, *67*, 390.
14. Mirjalili, B. F.; Bamoniri, A.; Akbari, A.; Taghavinia, N. *J. Iran. Chem. Soc.* **2011**, *8*, S129.
15. Biginelli, R. *Gazz. Chim. Ital.* **1893**, *23*, 360.
16. Nagarajaiah, H.; Mukhopadhyay, A.; Moorthy, J. N. *Tetrahedron Lett.* **2016**, *57*, 5135.
17. Alvim, H. G. O.; da Silva Junior, E. N.; Neto, B. A. D. *RSC Adv.* **2014**, *4*, 54282.
18. Sandhu, S.; Sandhu, J. S. *Arkivoc* **2012**, *66*.
19. Banik, B. K.; Reddy, A. T.; Datta, A.; Mukhopadhyay, C. *Tetrahedron Lett.* **2007**, *48*, 7392.
20. Li, J. T.; Han, J. F.; Yang, J. H.; Li, T. S. *Ultrason. Sonochem.* **2003**, *10*, 119.
21. Peng, J. J.; Deng, Y. Q. *Tetrahedron Lett.* **2001**, *42*, 5917.
22. Ma, Y.; Qian, C.; Wang, L.; Yang, M. *J. Org. Chem.* **2000**, *65*, 3864.

23. Tu, S. J.; Fang, F.; Miao, C. B.; Jiang, H.; Feng, Y. J.; Shi, D. Q.; Wang, X. S. *Tetrahedron Lett.* **2003**, *44*, 6153.
24. Sabitha, G.; Reddy, G. S. K. K.; Reddy, K. B.; Yadav, J. S. *Tetrahedron Lett.* **2003**, *44*, 6497.
25. Su, W. K.; Li, J. J.; Zheng, Z. G.; Shen, Y. C. *Tetrahedron Lett.* **2005**, *46*, 6037.
26. Hatamjafari, F. *Org. Chem. Int.* **2014**, *4*, 1.
27. Fu, N. Y.; Yuan, Y. F.; Pang, M. L.; Wang, J. T.; Peppe, C. J. *Organomet. Chem.* **2003**, *672*, 52.
28. Azizian, J.; Mohammadi, A. A.; Karimi, A. R.; Mohammadizadeh, M. R. *Appl. Catal., A Gen.* **2006**, *300*, 85.
29. Salehi, P.; Dabiri, M.; Zolfogol, M. A.; Fard, M. A. B. *Tetrahedron Lett.* **2003**, *44*, 2889.
30. Kumar, K. A.; Kasthuraiah, M.; Reddy, C. S.; Reddy, C. D. *Tetrahedron Lett.* **2001**, *42*, 7873.
31. Nandurkar, N. S.; Bhanushali, M. J.; Bhor, M. D.; Bhanage, B. M. *J. Mol. Catal. A: Chem.* **2007**, *271*, 14.
32. Tajbakhsh, M.; Mohajerani, B.; Heravi, M. M.; Ahmadi, A. N. *J. Mol. Catal. A: Chem.* **2005**, *236*, 216.
33. Ahmed, N.; van Lier, J. E. *Tetrahedron Lett.* **2007**, *48*, 5407.
34. Adib, M.; Ghanbari, K.; Mostofi, M.; Ganjali, M. R. *Molecules* **2006**, *11*, 649.
35. Abbasi, E.; Hatamjafari, F. *Orient. J. Chem.* **2013**, *29*, 731.
36. Gharib, A.; Jahangir, M.; Roshani, M.; Scheeren, J. W.; Moghadaszadeh, S.; Rezaee, A. *GUJ Sci.* **2012**, *25*, 823.
37. Patil, S.; Jadhav, S. D.; Deshmukh, M. B. *Arch. Appl. Sci. Res.* **2011**, *3*, 203.
38. Sheldon, R. A.; van Bekkum H. Eds.; *Fine Chemical through Heterogeneous Catalysis*. Wiley–VCH: Weinheim, Germany, 2002.
39. Cadman, J.; Zhou, S.; Chen, Y.; Li, Q. *J. Bionic Eng.* **2012**, *9*, 367.
40. Birchall, J. D.; Thomas, N. L. *J. Mater. Sci.* **1983**, *18*, 2081.
41. Gower, D.; Vincent, J. F. V. *Biomimetics*, **1996**, *4*, 37.
42. Sherrard, K. M. *Biol. Bull.* **2000**, *198*, 404.
43. Falini, G.; Fermani, S. *Tissue Eng.* **2004**, *10*, 1.
44. Ward, P. D.; Boletzky, S. V. *J. Mar. Biol. Assoc. U. K.* **1984**, *64*, 955.
45. Vogel, S. *J. Biosciences* **2006**, *31*, 309.
46. Cadman, J.; Chen, Y.; Zhou, Sh.; Li, Q. *Adv. Mater. Res.* **2010**, *123*, 295.
47. Jothi, N.; Kunthavai Nachiyar, R. *Global J. Biotechnol. Biochem.* **2013**, *8*, 33.
48. Ghodsinia, S. E.; Akhlaghinia, B. *RSC Adv.* **2015**, *5*, 49849.
49. Osada, M.; Miura, C.; Nakagawa, Y. S.; Kaihara, M.; Nikaido, M.; Totani, K. *Carbohydr. Polym.* **2013**, *92*, 1573.
50. Pearson, F. G.; Marchessault, R. H.; Liang, C. Y. *J. Polym. Sci.* **1960**, *43*, 101.
51. Khodja, I. A.; Boulcina, R.; Debaché, A. *Lett. Org. Chem.* **2015**, *12*, 77.
52. Mohamadpour, F.; Maghsoodlou, M. T.; Heydari, R.; Lashkari, M. *J. Iran. Chem. Soc.* **2016**, *13*, 1549.
53. Liu, Z.; Ma, R.; Cao, D.; Liu, C. *Molecules* **2016**, *21*, 462.
54. Fazaeli, R.; Aliyan, H.; Mohammadifar, F.; Zamani, A. A.; Bagi, M. J. *J. Korean. Chem. Soc.* **2011**, *55*, 666.
55. Dadhania, A. N.; Patel, V. K.; Raval, D. K. *J. Chem. Sci.* **2012**, *124*, 921.
56. (a) Foroughifar, N.; Mobinikhaledi, A.; Fathinejad, H. *Phosphorus, Sulfur Silicon Relat. Elem.* **2003**, *178*, 495. (b) Firdous, H.; Zafar, A. M.; Khan, M. N.; Parveen, N.; Munawar, A. M.; Khan, M. A. *Asian J. Chem.* **2015**, *27*, 455. (c) Salehi, H.; Guo, Q.X. *Synth. Commun.* **2004**, *34*, 171. (d) Nasr-Esfahani, M.; Montazerozohori, M.; Aghel-Mirrezaee, M.; Kashi, H. *J. Chil. Chem. Soc.* **2014**, *59*, 2311. (e) Ramos, L. M.; Guido, B. C.; Nobrega, C. C.; Correa, J. R.; Silva, R. G.; De Oliveira, H. C. B.; Gomes, A. F.; Gozzo, F. C.; Neto, B. A. D. *Chem. Eur. J.* **2013**, *19*, 4156.