

Chemical Constituents of *Psacalium sinuatum*

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Abstract. A chemical study of *Psacalium sinuatum* afforded the furanoeremophilane decompostine, the modified eremophilanes: cacalol, cacalol acetate, dehydrocacalol, maturinin, cacalohastin, dehydrocacalohastin, adenostin B, cacalone-*epi*-cacalone mixture, dimaturin, adenostylide, radulifolin F, and *epi*-radulifolin F, in addition to hyperin, sucrose and a mixture of sitosterol-stigmasterol. Extracts and isolated compounds were tested for anti-inflammatory activity. The hexanic extract of root showed a dose dependent activity (IC₅₀ 0.20 mg/ear).

Key words. *Psacalium*, Asteraceae, Senecioneae, Tussilaginatae, Modified Eremophilanes, Anti-inflammatory Activity.

Resumen. Por medio de un estudio químico de *Psacalium sinuatum* se aislaron el furanoeremofilano decompostina, los eremofilanos modificados: cacalol, acetato de cacalol, deshidrocacalol, maturinina, cacalohastina, deshidrocacalohastina, adenostina B, mezcla de cacalone-*epi*-cacalone, dimaturina, adenostilida, radulifolina F, y *epi*-radulifolina F, además de hiperina, sacarosa y mezcla de sitosterol-estigmasterol. Los extractos y los compuestos aislados fueron sometidos a pruebas de actividad anti-inflamatoria. El extracto hexánico de raíz presentó actividad dependiente de la dosis (IC₅₀ 0.20 mg/oreja).

Palabras clave: *Psacalium*, Asteraceae, Senecioneae, Tussilaginatae, eremofilanos modificados, actividad anti-inflamatoria.

Introduction

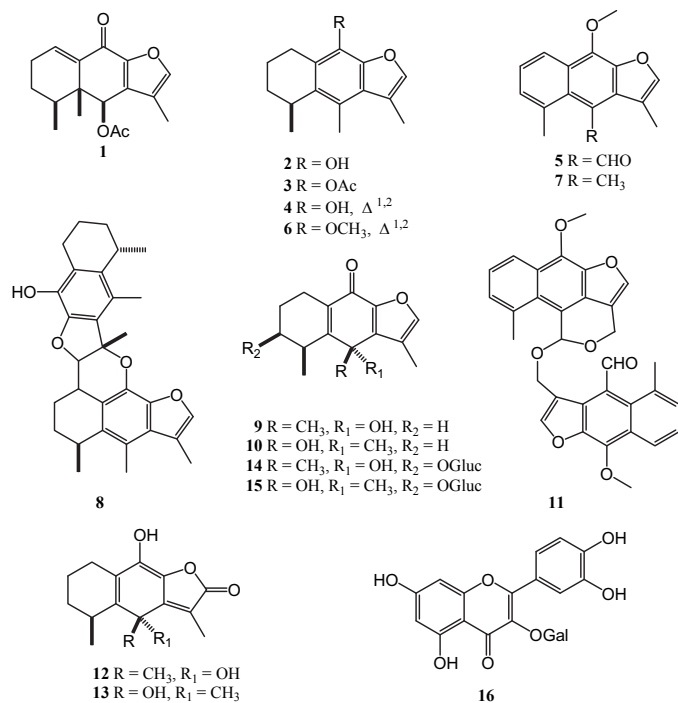
The genus *Psacalium* (Asteraceae, Senecioneae, Tussilaginatae) groups 40 species of perennial herbs disseminated from the south of the United States to Guatemala [1]. *P. sinuatum* is one of the six species included into the Matarique medicinal plant complex which is used in Mexico by the Tarahumara ethnica to cure diabetes, renal and hepatic problems [2]. So far, seven species of *Psacalium* have been chemically studied. Four of them: *P. decompositum*, *P. tussilaginoide*s (studied as *Cacalia decomposita* and *C. ampulacea*, respectively) [3-5], *P. radulifolium* [6] and *P. beamanii* [7] afforded cacalol derivatives and some eremophilanes mainly. The fifth, *P. paucicapitatum* [8] produced furanoeremophilanes and eremophilanolides, and the other two, *P. megaphyllum* [9] and *P. peltatum* (with only the aqueous extract studied) [10] yielded only a flavonoid glycoside and an ulopyranose, respectively. Several biological activities from species of *Psacalium* have been reported, thus, the hypoglycemic activity of the root water decoction of *P. decompositum* and the anti-hyperglycemic activity of some eremophilanolides isolated from this species is well documented [5, 11]. A hypoglycemic carbohydrate has been isolated from *P. peltatum* [10]. Cacalol has been detected as the major active compound in the antioxidant and antimicrobial evaluations of *P. radulifolium* [6, 12]. The anti-inflammatory activity of cacalol, cacalone [13], cacalohastin and radulifolin F [14] has also been reported. As continuation of our survey on Senecioneae we report the chemical composition of *P. sinuatum*, which to the best of our knowledge has no previous studies. An evaluation of the anti-inflammatory activity of extracts and isolated compounds on 12-O-tetradecanoylphorbol-13-acetate (TPA) model of induced acute inflammation is also included.

Results and Discussion

The chemical study of roots and aerial parts of *Psacalium sinuatum* afforded the furanoeremophilane decompostine (**1**) [7], the modified eremophilanes: cacalol (**2**) [15], cacalol acetate (**3**) [16], dehydrocacalol (**4**) [17], maturinin (**5**) [3], cacalohastin (**6**), dehydrocacalohastin (**7**) [18], adenostin B (**8**) [19], a 1:1 mixture of cacalone (**9**) and *epi*-cacalone (**10**) [5], dimaturin (**11**) [3], adenostylide (1:1 mixture of **12** and **13**) [19], radulifolin F (**14**), and *epi*-radulifolin F (**15**) [6], in addition to hyperin (**16**) [20], sucrose and a mixture of sitosterol-stigmasterol. Structures of the isolated compounds were determined by comparison of their physical constants and spectroscopic data with those reported in the literature.

Nearly all the compounds isolated from *P. sinuatum* are modified eremophilane derivatives oxygenated at positions 8 and 9 (**2-15**), which have also been reported as the more abundant metabolites in four of the six *Psacalium* species studied so far: *P. decompositum*, *P. tussilaginoide*s [3-5], *P. radulifolium* [6], and *P. beamanii* [7]. Pyrrolizidine alkaloids, also common in Senecioneae, have not been found in *Psacalium*, with the only exception of *P. decompositum* in whose aqueous extract these alkaloids were detected by TLC [5]. In summary, even though five out the seven species of *Psacalium* studied so far have shown a similar composition, at this point, it is not possible to establish a chemical profile of the genus with only 17.5% of the species studied.

On the other hand, the anti-inflammatory activity of the hexanic, acetic and methanolic extracts of roots and aerial parts, as well as that of the isolated compounds was tested on 12-O-tetradecanoylphorbol-13-acetate (TPA) model of induced acute inflammation [22] (Table 1). The hexanic extract of roots showed a maximal edema inhibition of 77.98% in a topical



administration of 1 mg/ear, this activity was dose dependent with a IC₅₀ of 0.2 mg/ear (Table 2). Indomethacin, the reference compound, produced 87.61% of edema inhibition at the same doses (1 mg/ear), but considering its IC₅₀ of 0.1 mg/ear, it was twice as active. The root extracts were always more active than the respective extracts of aerial parts; the activity in both, roots and aerial parts, decreases with the increase in the polarity of the extract (Table 1).

Among the tested compounds, the mixture of cacalone (**9**) and *epi*-cacalone (**10**) was more active than cacalol (**2**), in concordance with literature [13]. In the compounds with cacalol skeleton, the activity decreases from 38.61% in cacalol (**2**) to 31.14% in cacalol acetate (**3**) and to 10.84% in cacalohastine (**6**), suggesting that somehow the free hydroxy group is important in the edema inhibition. Moreover, the increase in activity to 59.40% observed in dehydrocacalol (**4**) may imply that planarity could also affect the inflammatory response, since dehydrocacalohastine (**7**) showed higher activity (35.80%) than cacalohastine (**6**) (10.84%). In the case of the mixture of cacalone (**9**) and *epi*-cacalone (**10**) the activity (50.33%) diminished with the presence of a glucosyloxy group in radulifolin F (**14**, 21.09%) and *epi*-radulifolin F (**15**, 13.74%).

Table 1. Effect of extracts and compounds isolated from *P. sinuatum* on TPA-induced mouse edema

Sample	Dose (mmol / ear)	Edema (mg)	Inhibition (%)
Hexanic extract of roots ^b	1.0 ^a	3.14 ± 0.31**	77.98**
Acetonic extract of roots ^c	1.0 ^a	5.03 ± 2.38*	56.73*
Methanolic extract of roots ^c	1.0 ^a	8.53 ± 1.98	26.65
Hexanic extract of aerial parts ^b	1.0 ^a	4.93 ± 1.74**	59.40**
Acetonic extract of aerial parts ^c	1.0 ^a	8.80 ± 1.21**	27.54**
Methanolic extract of aerial parts ^c	1.0 ^a	10.57 ± 0.07*	21.87*
Decompostin (1) ^b	1.0	7.67 ± 1.91*	36.90*
Cacalol (2) ^d	1.0	9.43 ± 0.29*	38.61*
Cacalol acetate (3) ^b	1.0	8.37 ± 1.88*	31.14*
Dehydrocacalol (4) ^b	1.0	4.93 ± 1.23**	59.40**
Maturinin (5) ^b	1.0	6.53 ± 1.19**	46.23**
Cacalohastine (6) ^b	1.0	10.83 ± 1.71	10.84
Dehydrocacalohastine (7) ^b	1.0	7.80 ± 1.82*	35.80*
Adenostin B (8) ^b	1.0	6.10 ± 2.09**	49.79**
Cacalone/ <i>epi</i> -cacalone (9/10 , 1:1) ^d	1.0	7.63 ± 1.62**	50.33**
Dimaturin (11) ^b	1.0	5.83 ± 1.30**	52.02**
Adenostylide (12/13 1:1) ^b	1.0	5.33 ± 2.12**	56.10**
Radulifolin F (14) ^e	1.0	10.27 ± 0.62*	21.09*
<i>Epi</i> -radulifolin F (15) ^e	1.0	11.67 ± 0.35	13.74
Hyperin (16) ^e	1.0	13.10 ± 0.71	3.14
Indomethacin ^f	1.0 ^a	2.06 ± 0.30**	87.61**
Indomethacin ^f	1.0	1.99 ± 0.69*	83.73*

Each value represents the mean of three animals ± standard error. ^aDose mg/ear. Control: ^bacetone-CH₂Cl₂ 1:1, 12.15 ± 0.98; ^cmethanol, 11.63 ± 0.24; ^dacetone-CH₂Cl₂ 1:1, 15.37 ± 1.85; ^eacetone, 13.53 ± 104; ^fethanol-acetone 1:1, 15.00 ± 0.47. Results were analysed by the *t* Student test. **p* ≤ 0.05. ***p* ≤ 0.01.

Table 2. Dose response evaluation of the hexanic extract of roots of *P. sinuatum*

Sample	Dose (mg/ear)	Edema (mg)	Inhibition (%)	IC ₅₀
Control		14.26 ± 0.99		
Hexanic extract of roots	0.031	12.34 ± 1.36	13.46	0.20 mg / ear r=0.97
	0.1	8.38 ± 1.07**	41.23**	
	0.31	6.86 ± 1.33**	51.89**	
	1.0	3.14 ± 0.31**	77.98**	
Indomethacin	0.036	11.83 ± 0.70*	21.11*	0.10 mg / ear r=0.99
	0.11	6.82 ± 1.27**	54.53**	
	0.357	1.99 ± 0.68**	83.73**	

Each value represents the mean of five animals ± standard error. Results were analysed by the analysis of variance ANOVA followed by Dunnett test. *p ≤ 0.05. **p ≤ 0.01

General Experimental Procedures

Melting points were determined on a Fisher Jones melting point apparatus and are uncorrected. IR spectra were recorded on a Nicolet Magna-IR 750 spectrometer. EIMS data were determined on a JEOL JMS-AX505HA mass spectrometer at 70 eV. ¹H NMR and ¹³C NMR data were obtained on a Varian Unity 300 or on a Varian Unity Plus 500 instrument. Chemical shifts were referred to TMS (δ 0). Column chromatography (VCCs) was performed under vacuum using Silica gel 60 G (Merck, Darmstadt, Germany). Flash column chromatography (FCC) was performed on silica gel 60 (230-400 Macherey-Nagel). TLC was carried out on Silica gel 60 GF₂₅₄ and preparative TLC on Silica gel GF₂₅₄ (Macherey-Nagel), layer thickness 2.0 mm.

Plant Material

Psacalium sinuatum (Cerv.) H. Rob. & Bretell was collected 6 km northeast of Canoas, Nayarit, México, in September 2007. A voucher specimen was deposited at the Herbarium del Instituto de Biología, UNAM, México (MEXU 1213451).

Extraction and Isolation

Dried and ground roots (130 g) and aerial parts (276 g) of *P. sinuatum* were separately and successively extracted with hexane, acetone and methanol. Solvents were removed at reduced pressure to obtain the respective extracts which gave negative the Dragendorff's alkaloids test. The hexanic extract of roots (6.0 g) was submitted to vacuum column chromatography (VCC) using hexane-EtOAc mixtures as gradient elution system to afford fractions A-C. Fraction A (1.8 g) was purified by a VCC eluted with hexane-EtOAc mixtures of increasing polarity to obtain from fractions eluted with hexane: cacalo-

hastine (**6**, 48 mg, 0.04%), mp 82-84° (lit. 84.0-85.5°) [18], dehydrocacalohastine (**7**, 30 mg, 0.02%), mp 73-75° (lit. 71.0-77.3°) [18], and cacalol acetate (**3**, 23 mg, 0.02%), mp 101-102° (lit. 105°) [16]. The same column produced from fractions eluted with hexane-EtOAc 49:1, adenostin B (**8**, 80 mg, 0.06%), mp 208-209° (lit. 202-205°) [19] and those fractions eluted with hexane-EtOAc 19:1 afforded a 1:1 mixture of cacalone (**9**) and *epi*-cacalone (**10**) (302 mg, 0.23%), mp 115-117° (lit. 115-118°) [5, 12]. Fraction B (2.5 g) was submitted to flash column chromatography (FCC) eluted with hexane-EtOAc 49:1 to obtain cacalol (**2**, 180 mg, 0.14%), mp 90-91° (lit. 90-92°) [12, 15] and maturinin (**5**, 10 mg, 0.01%), mp 95-96° (lit. 95-96°) [3]. From fraction C decompostin (**1**, 330 mg, 0.25%), mp 192-194° (lit. 195-196°) [7] was isolated. Purification of its mother liquors (1.8 g) by FCC eluted with hexane-EtOAc 9:1 produced fractions D and E. Fraction D (350 mg) was submitted to FCC eluted with hexane-acetone 19:1 to obtain a 1:1 mixture of **9** and **10** (130 mg, 0.1%). Purification of fraction E (220 mg) by FCC eluted with hexane-EtOAc 19:1 produced dimaturin (**11**, 30 mg, 0.02%), mp 199-201° (lit. 201-203°) [3], and **1** (50 mg, 0.04%). The acetonic extract of roots (3.0 g) afforded fractions F and G by VCC using hexane-EtOAc as elution solvent. Fraction F (300 mg) was purified by FCC eluted with hexane-EtOAc 49:1 to obtain **3** (40 mg, 0.03%) and **2** (83 mg, 0.06%). From fraction G (500 mg), compound **3** (10 mg, 0.01%) and adenostylide (1:1 mixture of **12** and **13**, 16 mg, 0.01%), mp 138-140° (lit. 140-141°) [19] were isolated by FCC eluted with hexane-EtOAc 19:1. The methanolic extract of roots (8.0 g) was purified by VCC using EtOAc-MeOH mixtures of increasing polarity. Fractions obtained with EtOAc-MeOH 49:1 (600 mg) were further purified by VCC eluted with CH₂Cl₂-MeOH 85:15 to afford fraction H (98 mg) which was purified by preparative TLC eluted three times with CH₂Cl₂-MeOH 9:1 to produce radulifolin F (**14**, 13 mg, 0.01%, colorless oil) and *epi*-radulifolin F (**15**, 11 mg, 0.01%, colorless oil) [6]. The hexanic extract of the aerial parts (5.0 g) was purified by two

successive chromatographies using hexane-EtOAc mixtures as elution solvents to produce **2** (85 mg, 0.03%), **3** (42 mg, 0.02%), dehydrocactalol (**4**, 30 mg, 0.01%), mp 120-122° (lit. 124-125°) [17] and 180 mg (0.07%) of a mixture sitosterol-stigmasterol. Purification of the acetonic extract of aerial parts (8.5 g) by VCC using hexane-EtOAc gradient mixtures, followed by a FCC eluted with hexane-EtOAc 49:1 led to the isolation of **1** (80 mg 0.03%), **2** (60 mg, 0.02%) and **3** (90 mg, 0.04%). The methanolic extract of aerial parts (24 g) was subjected to VCC eluted with EtOAc-MeOH in increasing gradient. Fractions eluted with EtOAc-MeOH 8:2 (12 g) were purified successively by VCC eluted with CH₂Cl₂-MeOH 9:1 and a Sephadex LH-20 column eluted with MeOH-H₂O 9:1 to produce hyperin (**16**, 450 mg, 0.16%), mp 243-245° (lit. 238-240°) [20, 21] and sucrose 1.5 g (0.54%).

Evaluation of the anti-inflammatory activity

Animals: Male NIH mice weighing 25-30 g were maintained in standard laboratory conditions in the animal house (temperature 27 ± 1 °C) in a 12/12 h light-dark cycle, according with the Mexican official norm MON-062-Z00-1999. They were fed laboratory diet and water *ad libitum*.

TPA-induced edema model

The TPA-induced ear edema assay in mice was performed as previously reported [22]. A solution of TPA (2.5 µg) in EtOH (10 µl) was applied topically to both faces (5 ml each face) of the right ear of the mice, 10 min after solutions of the test substances in their respective solvents (Table 1) were applied (10 µl each face). The left ear received ethanol (10 µl) first, and 20 ml of the respective solvent subsequently. Four hours later the mice were killed by cervical dislocation. A 7 mm diameter plug was removed from each ear. The swelling was assessed as the difference in weight between the left and the right ear. Control animals received the correspondent solvent in each case. Edema inhibition (EI %) was calculated by the equation: $EI = 100 - (B \times 100/A)$, where A is the edema induced by TPA alone and B is the edema induced by TPA plus sample. Indomethacin was used as the reference compound (Tables 1, 2)

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References

- Barkley, T. M.; Clark, B. L.; Fuston, A. M. *Compositae: Systematics*. Proceedings of the International Compositae Conference, Kew, 1994. Hind, D. J. N., Ed. Royal Botanical Gardens Kew, **1996**, Vol 1, pp. 613-620.
- Linares, E.; Bye, R. *J. Ethnopharmacol.* **1987**, *19*, 153-186.
- Correa, J.; Romo, J. *Tetrahedron* **1966**, *22*, 685-691.
- Joseph-Nathan, P.; Negrete, M. C.; González, M. P. *Phytochemistry* **1970**, *9*, 1623-1628.
- Inman, W. D.; Luo, J.; Joload, S. D.; King, S. R.; Cooper, R. J. *J. Nat. Prod.* **1999**, *62*, 1088-1092.
- Garduño-Ramírez, M. L.; Delgado, G. *Rev. Soc. Quím. Méx.* **2003**, *47*, 160-166.
- Pérez-Castorena, A. L.; Arciniegas, A.; Villaseñor, J. L.; Romo de Vivar, A. *Rev. Soc. Quím. Méx.* **2004**, *48*, 21-23.
- Burgueño-Tapia, E.; Hernández-Carlos, B.; Joseph-Nathan, P. *J. Mol. Struct.* **2006**, *825*, 115-123.
- Pérez-Castorena, A. L.; Castro, A.; Romo de Vivar, A. *Phytochemistry* **1997**, *46*, 1297-1299.
- Contreras, C.; Román, R.; Pérez, C.; Alarcón, F.; Zavala, M.; Pérez, S. *Chem. Pharm. Bull.* **2005**, *53*, 1408-1410.
- Alarcón-Aguilar, F. L.; Jiménez-Estrada, M.; Reyes-Chilpa, R.; González-Paredes, B.; Contreras-Weber, C. C.; Román-Ramos, R. *J. Ethnopharmacol.* **2000**, *69*, 207-215.
- Garduño-Ramírez, M. L.; Trejo, A.; Navarro, V.; Bye, R.; Linares, E.; Delgado, G. *J. Nat. Prod.* **2001**, *64*, 432-435.
- Jiménez-Estrada, M.; Reyes Chilpa, R.; Ramírez Apan, T.; Lledias, F.; Hansberg, W.; Arrieta, D.; Alarcón Aguilar, F. J. *J. Ethnopharmacol.* **2006**, *105*, 34-38.
- Acevedo-Quiroz, N.; Domínguez-Villegas, V.; Garduño-Ramírez, M. L. *Nat. Prod. Comm.* **2008**, *3*, 313-317.
- Kedrowski, B. L.; Hoppe, R. W. *J. Org. Chem.* **2008**, *73*, 5177-5179.
- Bohlmann, F.; Zdero, C. *Phytochemistry* **1978**, *17*, 1135-1153.
- Kakisawa, H.; Inouye, I. *Tetrahedron Lett.* **1969**, *24*, 1929-1932.
- Hayashi, K.; Nakamura, H.; Mitsuhashi, H.; *Phytochemistry* **1973**, *12*, 2931-2933.
- Kuroyanagi, M.; Naito, H.; Noro, T.; Ueno, A.; Fukushima, S. *Chem. Pharm. Bull.* **1985**, *33*, 4792-4797.
- Markham, K. R.; Ternai, B.; Stanley, R.; Geiger, H.; Mabry, T. J. *Tetrahedron* **1978**, *34*, 1389-1397.
- Rizk, A. M. *Planta Med.* **1979**, *36*, 189-190.
- Pérez-Castorena, A. L.; Arciniegas, A.; Ramírez-Apan, M. T.; Villaseñor J. L.; Romo de Vivar, A. *Planta Med.* **2002**, *68*, 645-647.